

Biomimetic Lipid Membranes for the Study of Membrane Protein Structure

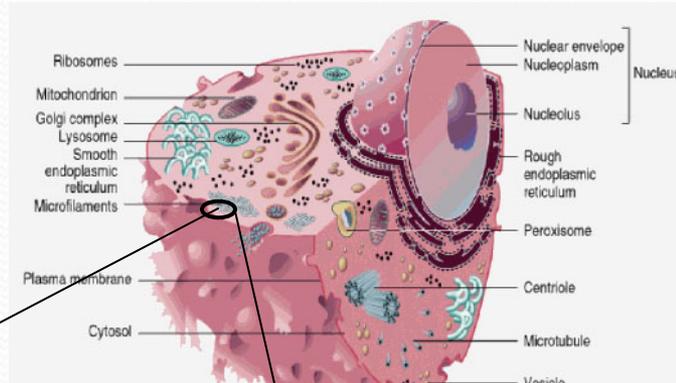
Emily Hoff, University of Southern Mississippi

SURF 2009

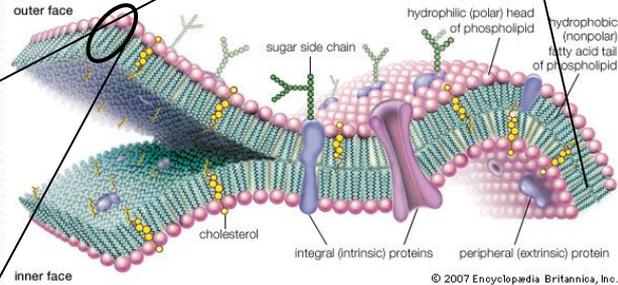


- ◎ **Goal**: Prepare model oriented lipid bilayers with incorporated membrane proteins to be studied with x-ray and neutron diffraction.
- ◎ **Part I**: Study lipid membranes and the effect of charge and pH on sample alignment and phase behavior.
- ◎ **Part II**: Study how the antimicrobial peptide, Gramicidin A, resides in the lipid membranes.

Cells



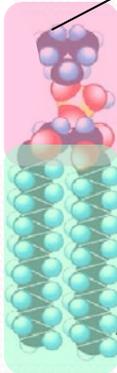
Membranes



Lipids

polar headgroup

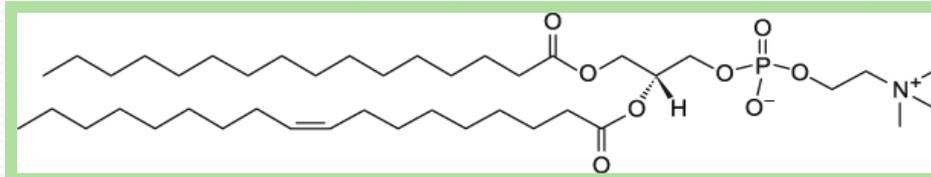
oily chains



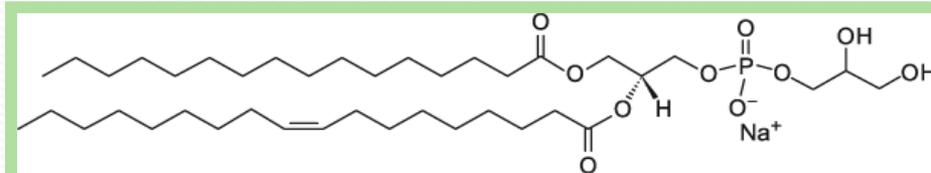
© 2007 Encyclopædia Britannica, Inc.

Comparison of Lipid Structures

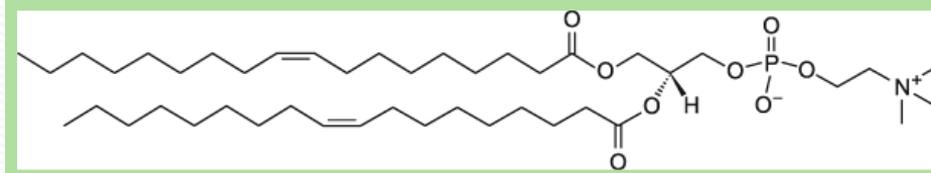
POPC



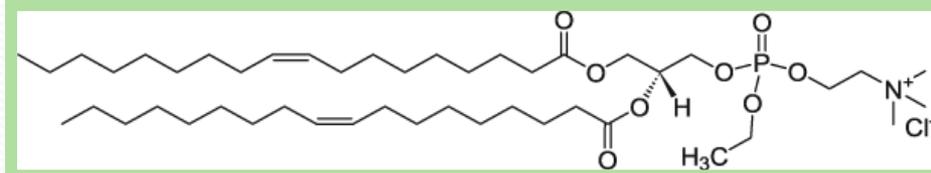
POPG



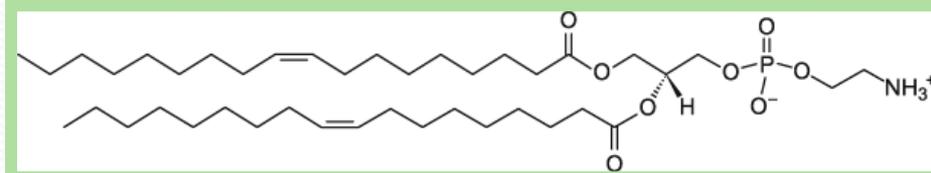
DOPC



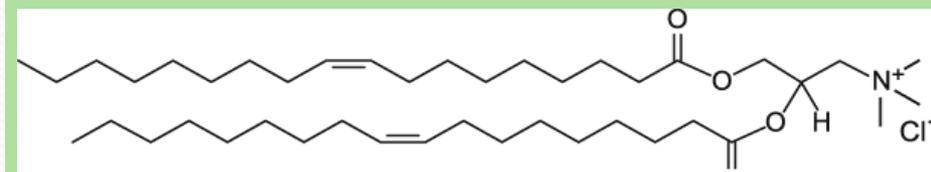
EDOPC



DOPE



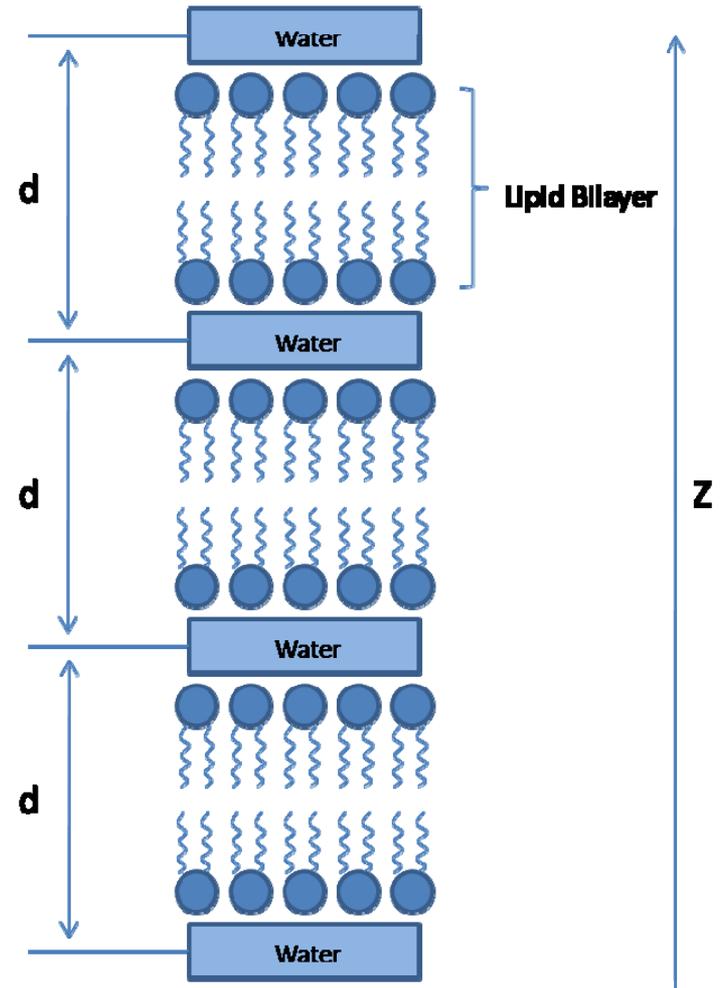
DOTAP



Introduction: Lipid Structure



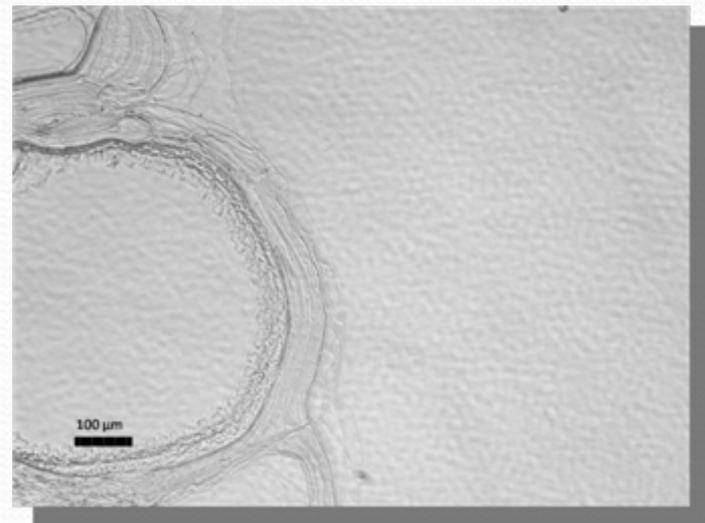
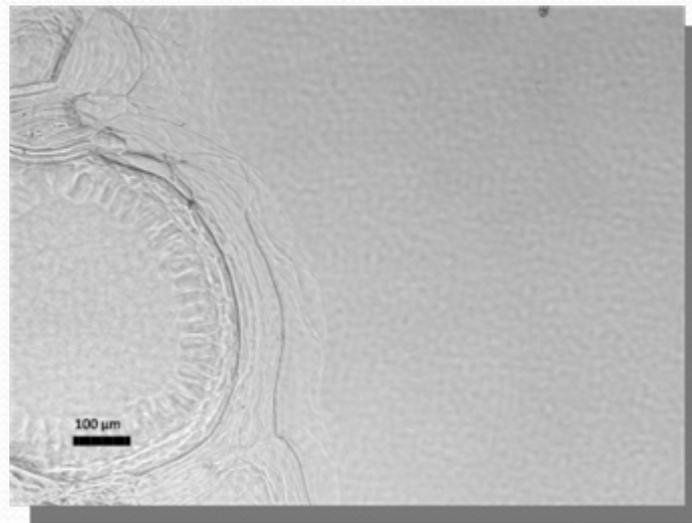
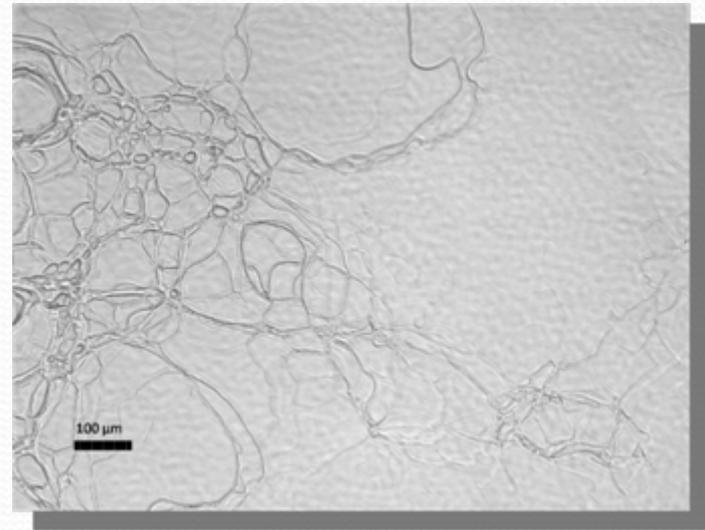
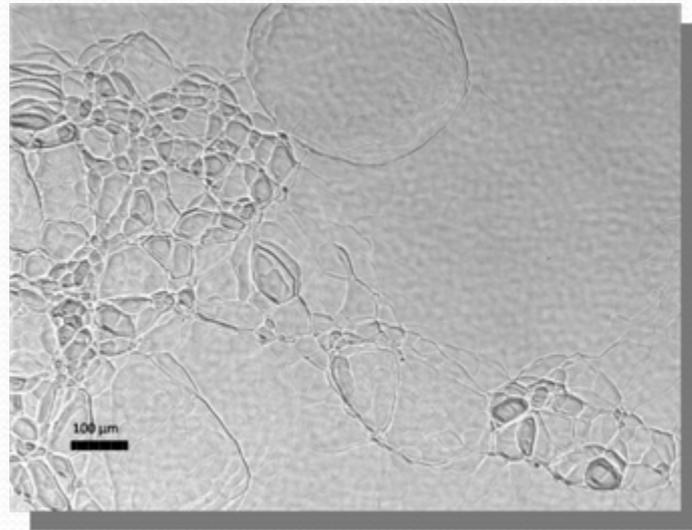
Lipid Multilayer



Microscope: PCPG (80:20)

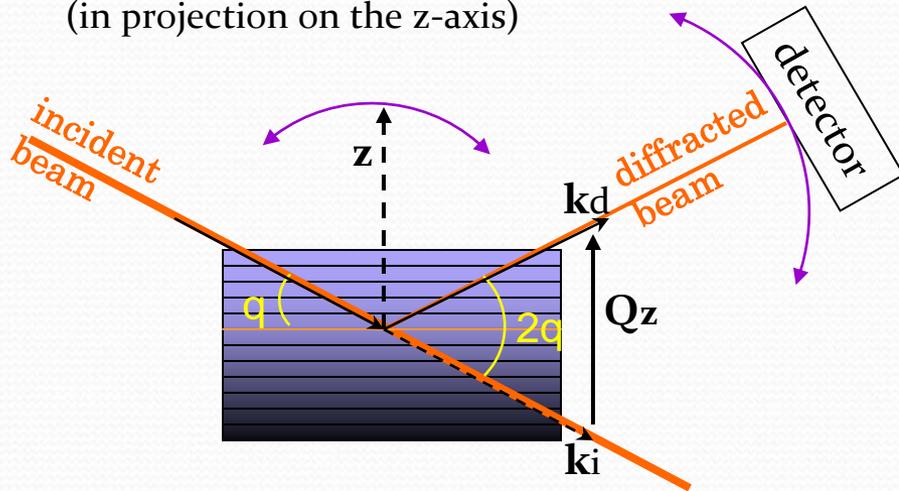
Before Hydration

After Hydration

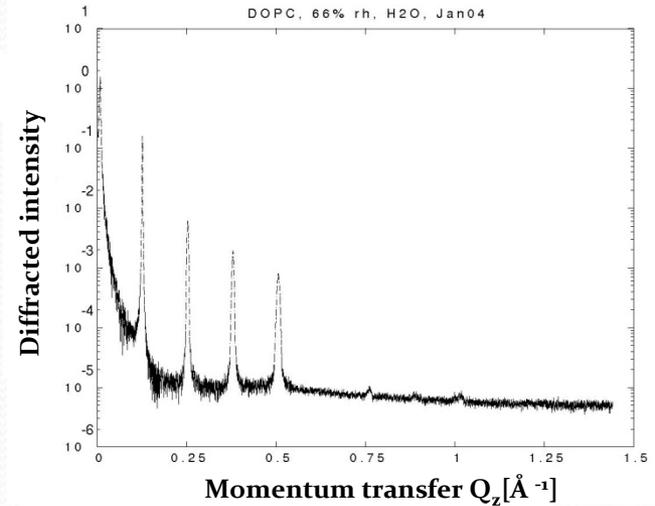


Measuring diffraction from oriented lipid multilayers

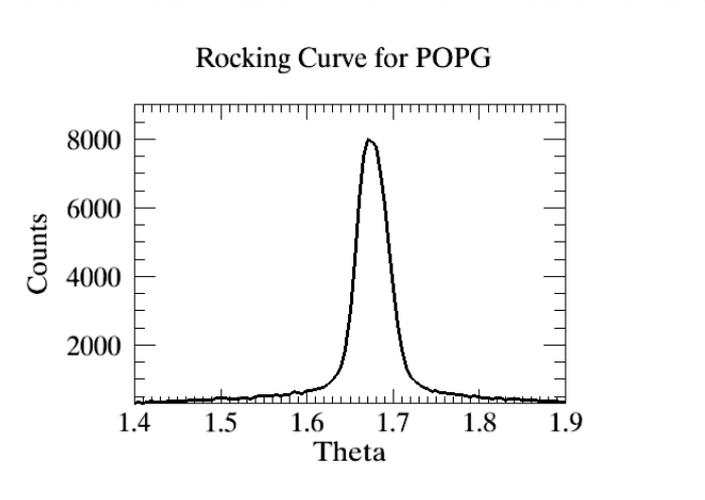
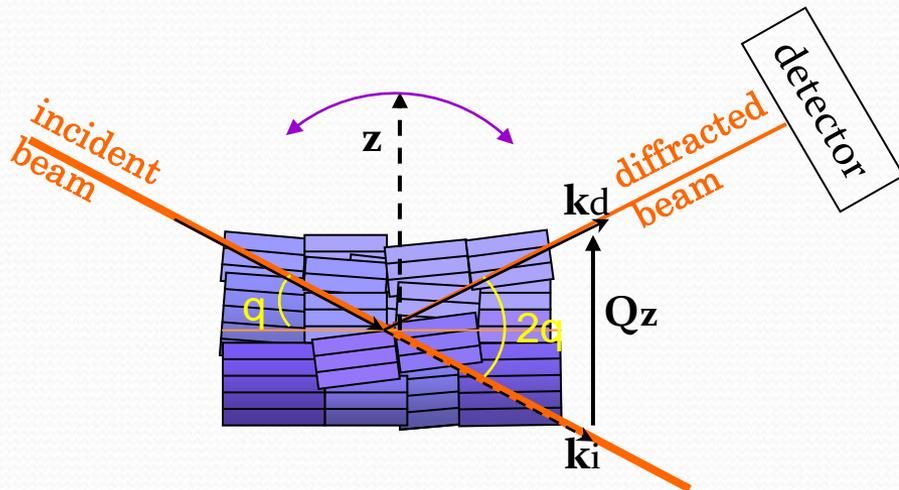
- θ - 2θ scans for determining lipid membrane structure (in projection on the z-axis)



$$Q_z = \frac{4\pi}{\lambda} \sin(\theta) = n \frac{2\pi}{d}$$



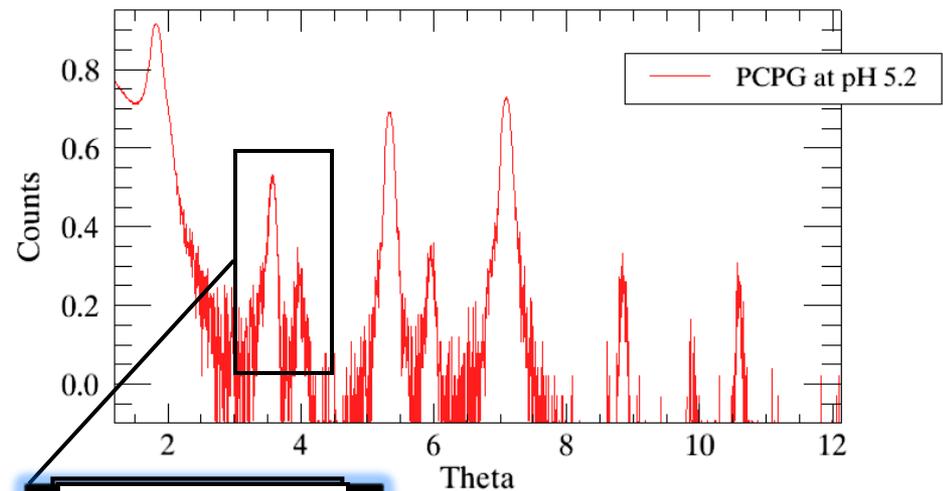
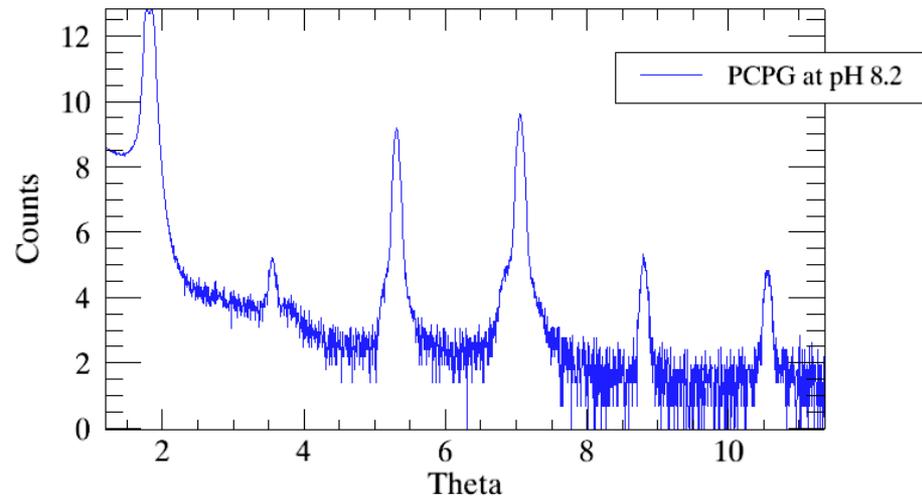
- θ scans for determining the degree of orientation



pH Effect on PG

Basic Environment: pH 8.2

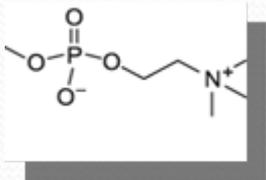
Acidic Environment: pH 5.2



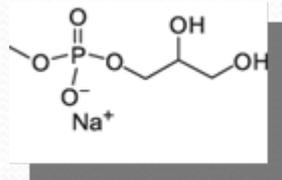
Additional
Diffraction
Peaks

$$Q_z = \frac{4\pi}{\lambda} \sin(\theta) = n \frac{2\pi}{d}$$

POPC



POPG



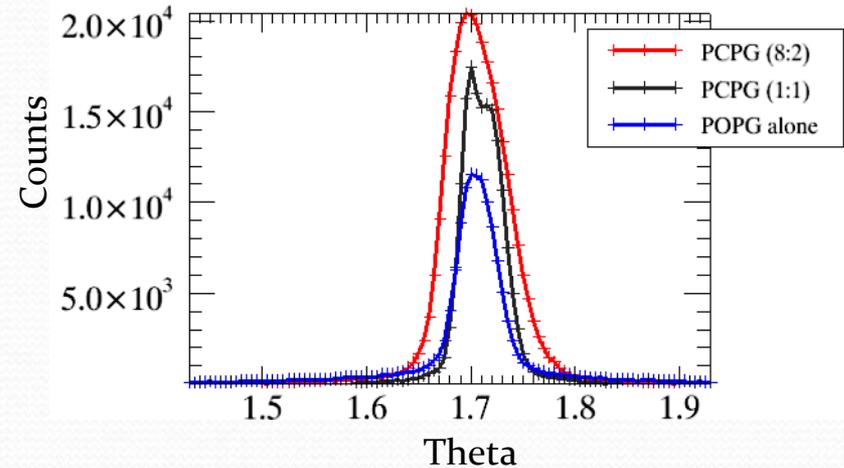
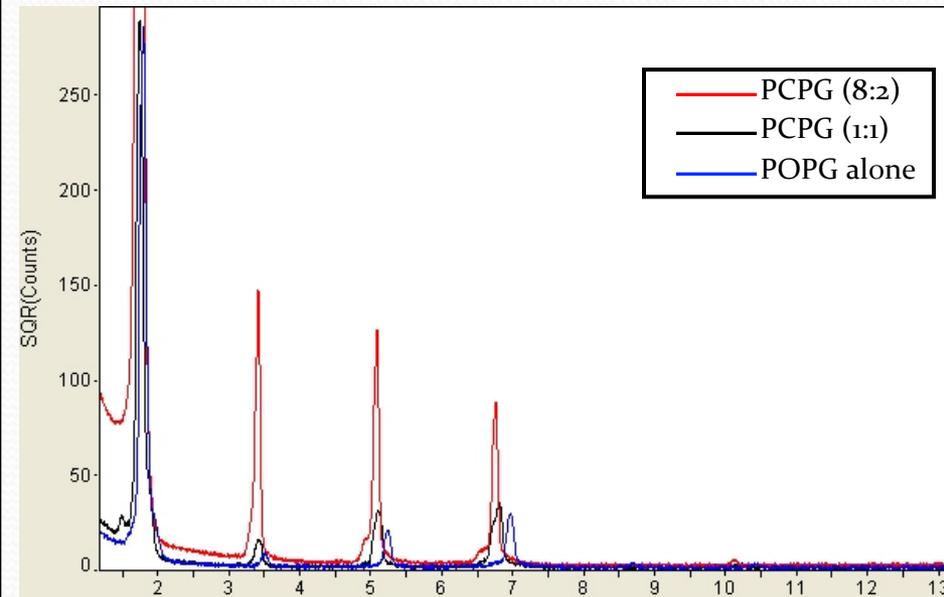
Phase separation of POPG from POPC in a acidic environment.

X-Ray Diffraction Results

PCPG(1:1) vs. PCPG(8:2) vs.
POPG alone

Rocking Curve

Rocking Curves of Lipids with Increasing PG content



Repeat
Distances →

PCPG (8:2):	52.6 +/- 0.07 Å
PCPG (1:1):	52.1 +/- 0.05 Å
POPG alone:	50.7 +/- 0.05 Å

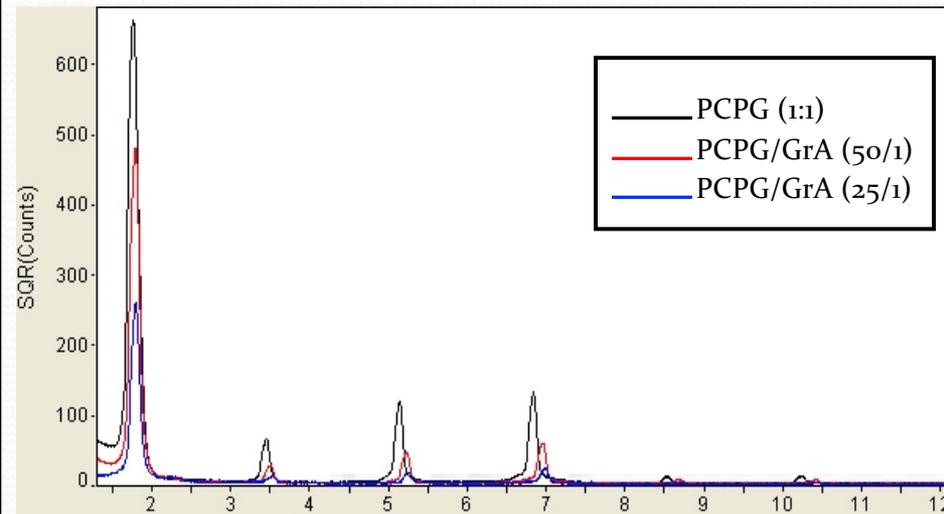
FWHM →

PCPG (8:2):	0.066 +/- 0.003
PCPG (1:1):	0.044 +/- 0.004
POPG:	0.041 +/- 0.001

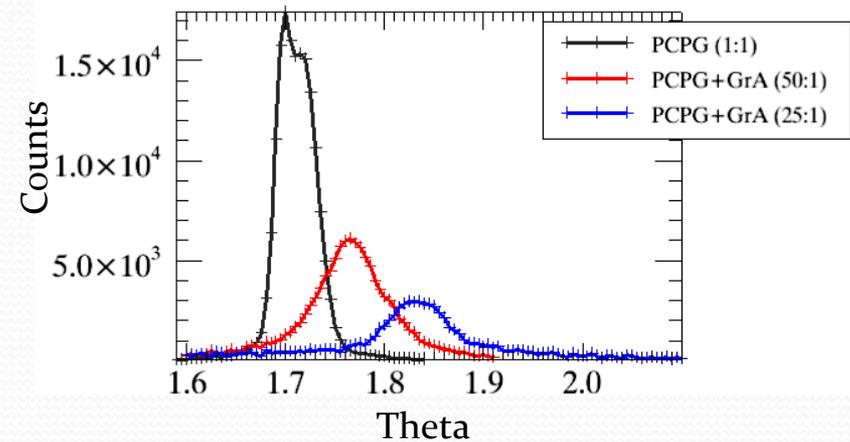
X-Ray Diffraction Results

PCPG/GrA(50/1) vs.
PCPG/GrA(25/1) vs. PCPG(1:1)

Rocking Curve



Rocking Curves of Lipids with Increasing GrA Content



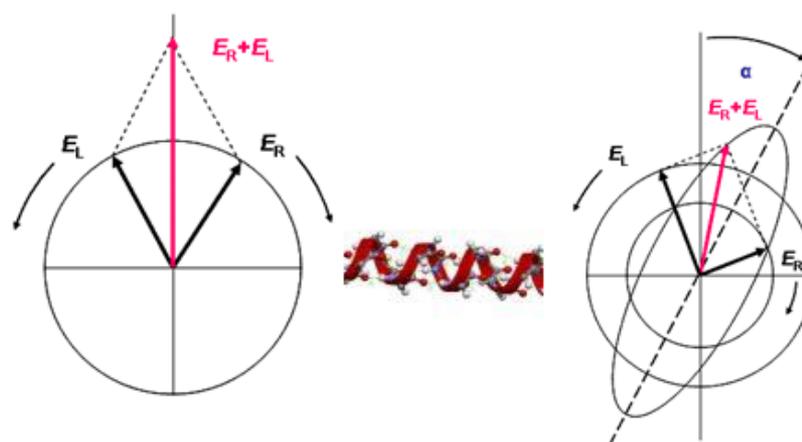
Repeat
Distances →

PCPG (1:1):	52.1 +/- 0.05 Å
PCPG+GrA (50:1):	49.2 +/- 0.05 Å
PCPG+GrA (25:1):	49.0 +/- 0.05

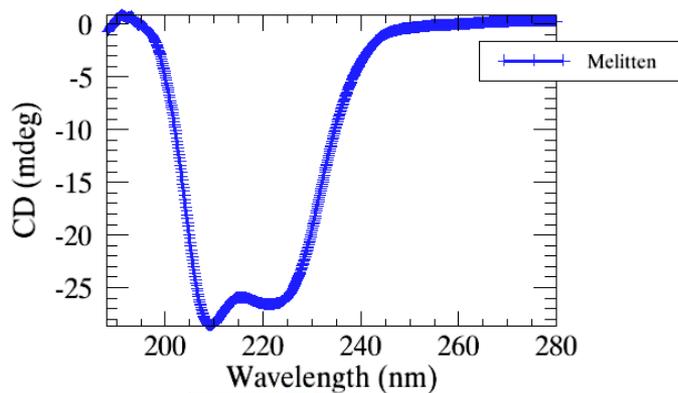
FWHM →

PCPG :	0.044 +/- 0.004
PCPG+GrA (50:1) :	0.072 +/- 0.003
PCPG+GrA (25:1) :	0.076 +/- 0.003

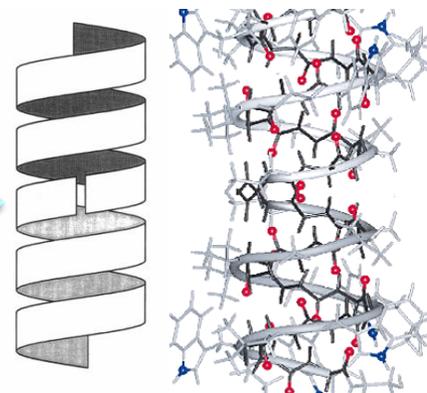
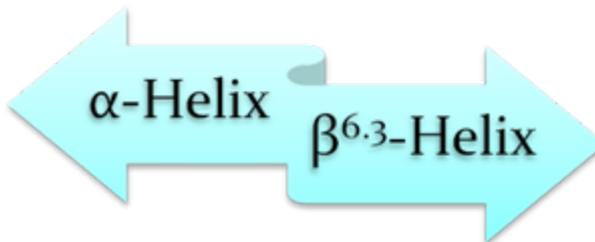
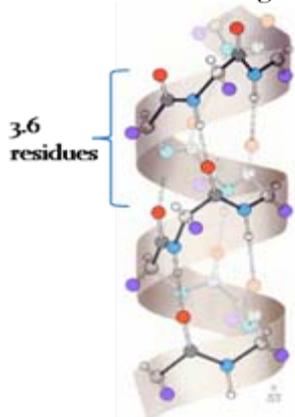
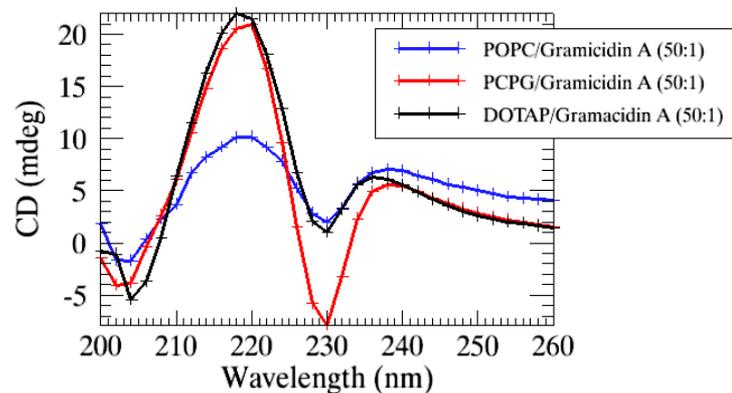
Circular Dichroism



CD Signal for an Alpha Helix Configuration



Gramicidin A in POPC, PCPG (1:1), and DOTAP



Neutron Diffraction



Advanced Neutron Diffractometer/Reflectometer
NIST Center for Neutron Research

What we measure

Intensity(n) = $1/n^2 N^2 |F(n)|^2$
for large number of bilayers, N

Fourier Sum

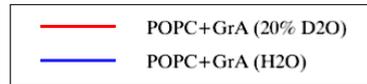
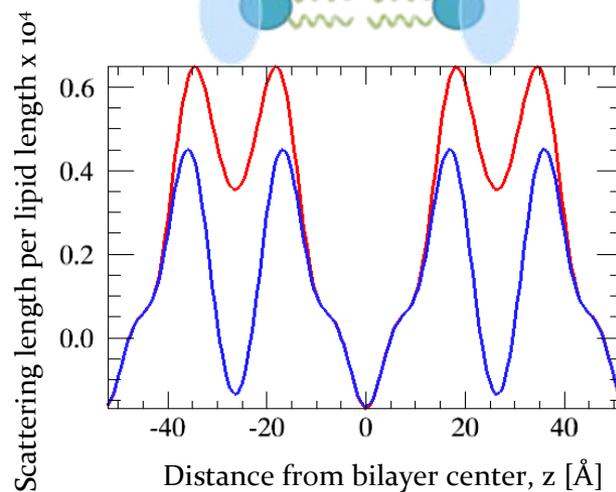
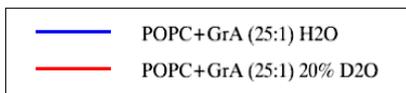
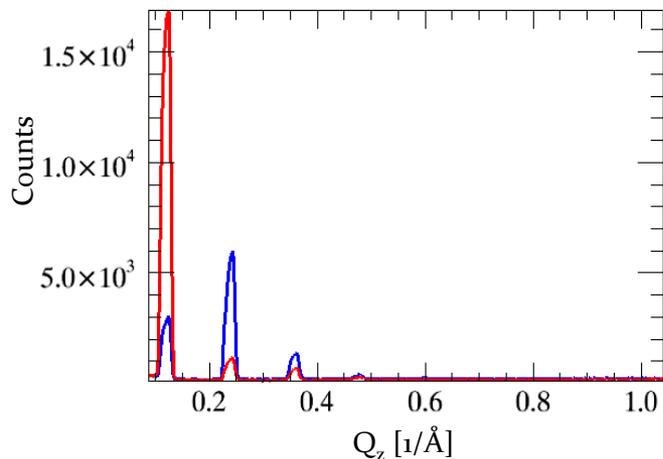
What we want to determine

$$\rho(z) = \frac{1}{d} \sum_n |F(n)| \cos\left(\frac{2\pi n z}{d}\right)$$

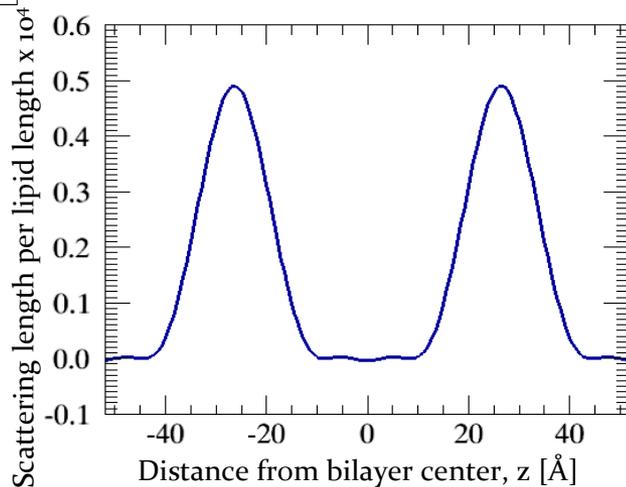
SLD Profiles for POPC+GrA in 20% D2O and H2O



POPC+Gramicidin A (25:1) (H2O vs. 20% D2O)



Water Distribution for POPC+GrA

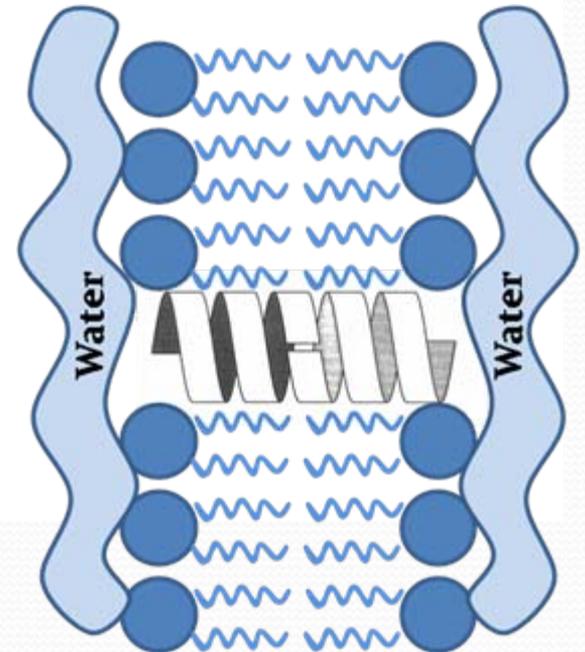
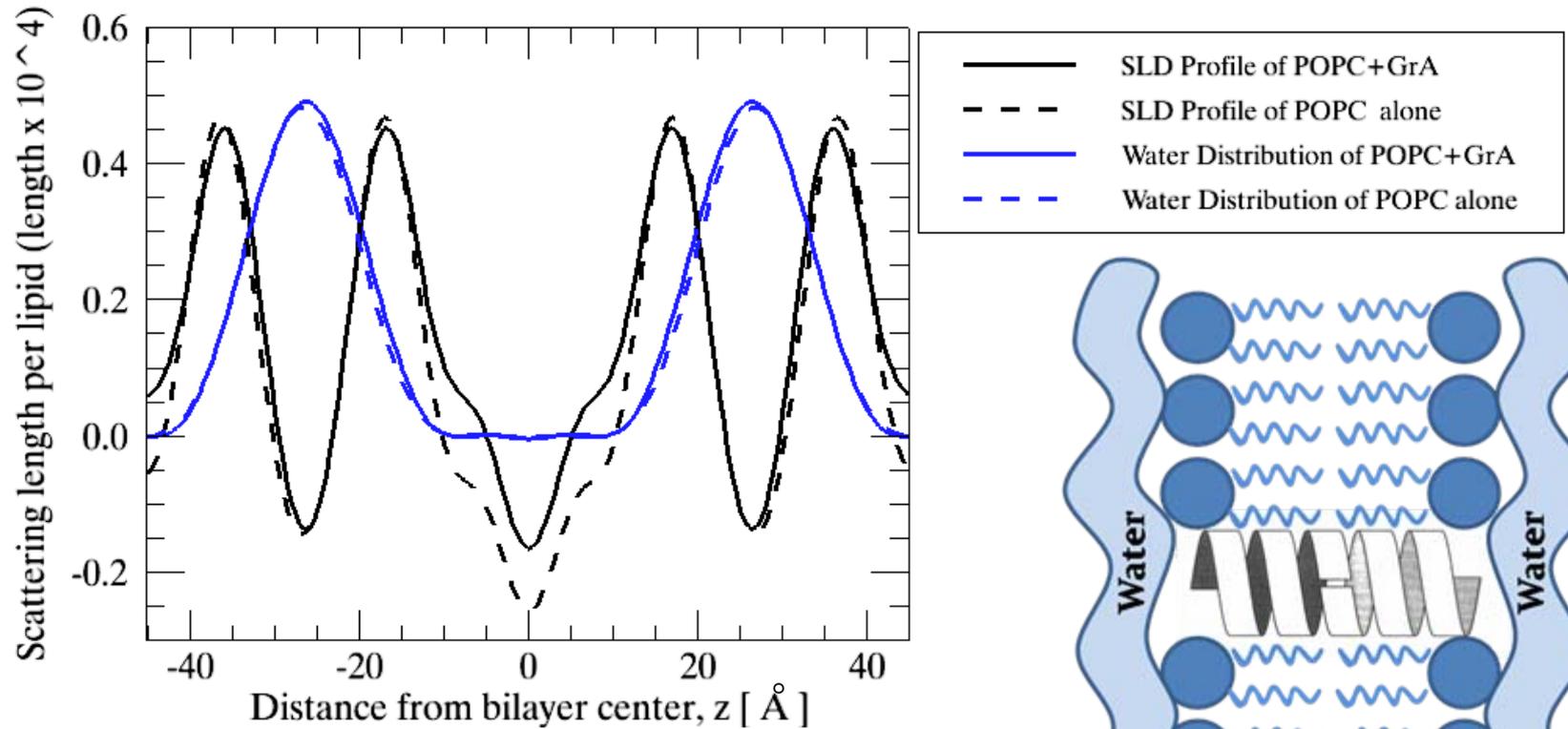


$$\rho(z)_D - \rho(z)_H$$

**Neutron
Diffraction
Data**

Neutron Diffraction Data

SLD Profiles and Water Distributions for POPC+GrA and POPC alone



$d = 53.5 \pm 0.08 \text{ \AA}$ (POPC alone)
 $d = 52.8 \pm 0.05 \text{ \AA}$ (POPC+GrA (25:1))

Conclusions

- ⦿ Improve on sample quality by hydration and the amount of charged lipid.
- ⦿ Solvent pH plays an important role in the structure and properties of a lipid membrane.
- ⦿ Neutron Diffraction and CD data indicate that Gramicidin A is incorporated into the membrane.
 - Suggests transmembrane, channel forming conformation.

Acknowledgements

- NIST and SURF Directors
- NCNR
- NSF and CHRNS
- NIH, Dr. Kenton Swartz's Lab
- Ella Mihailescu
- Dr. Julie Borchers
- University of Southern Mississippi



NCNR SURF

References:

1. Campbell, Wilbur H. Alpha Helix. Digital image. *BL/CH401 Lecture 8: Protein Secondary Structure Section E*. 1995. Web. 30 July 2009. <http://www.bio.mtu.edu/campbell/401lec8c.html>
2. Cross, T. A., A. Arseniev, B. A. Cornell, J. H. Davis, J. A. Killian, R. E. Koeppe, II, L. K. Nicholson, F. Serapovic, and B. A. Wallace. Single-Strand Conformation of Gramicidin. Digital image. *Nature Structural and Molecular Biology: Gramicidin Channel Controversy? Revisited*. Nature Publishing Group, 1999. Web. 30 July 2009. http://www.nature.com/nsmb/journal/v6/n7/full/nsbo799_610.html
3. Girshman, Jeffery, Denise V. Greathouse, Roger E. Koeppe,II, and Olaf S. Anderson. Beta 6.3 Helical Dimer. Digital image. *Gramicidin Channels in Phospholipid Bilayers with Unsaturated*. Biophysical Journal. Web. 30 July 2009. <http://www.pubmedcentral.nih.gov/picrender.fcgi?artid=1181031&blobtype=pdf>
4. Wang, Jane. Phospholipid and Lipid Bilayer. Digital image. *BioTeach*. Web. 30 July 2009. <http://www.bioteach.ubc.ca/Bio-industry/Inex/>