

## What Can be Measured by SANS and Reflectometry?

**Charles Glinka** 

**Reflectometry from Submicron Structures** NIST Summer School on SANS and

June 3-7, 2002







Incident neutron wave vector,  $\left|\vec{k}_{i}\right| = \frac{2\pi}{\lambda_{i}}$ 







**Center for Neutron Research** 

 $2\theta$  $d \approx \frac{2\pi}{0}$ , for small scattering angles,  $d \approx \frac{\lambda}{2\theta}$ 

In general, diffraction (SANS or NR) probes length scale

or 
$$d = \frac{\lambda}{2\sin\theta} = \frac{2\pi}{\left(\frac{4\pi}{\lambda}\right)\sin\theta} = \frac{2\pi}{Q}$$

Recall Bragg's Law 
$$\rightarrow \lambda = 2d \sin \theta$$
  
or  $d = \frac{\lambda}{2\sin\theta} = \frac{2\pi}{\left(\frac{4\pi}{2}\right)_{\sin\theta}} = \frac{2\pi}{Q}$ 

Secall Bragg's Law 
$$\rightarrow \lambda = \angle u$$
 SII  
or  $d = \frac{\lambda}{2\sin\theta} = \frac{2\pi}{(4\pi) \cdot \beta} = \frac{2\pi}{0}$ 

ecall Bragg's Law 
$$\longrightarrow \lambda = 2d \sin \beta$$

$$k_{i} \xrightarrow{2\theta} Q = \frac{Q}{\lambda} \operatorname{sin} \theta$$

$$Q = \left(\frac{4\pi}{\lambda}\right) \operatorname{sir}$$

$$|\vec{Q}| = 2k\sin\theta$$
  
 $Q = \left(\frac{4\pi}{\lambda}\right)\sin\theta$ 

Kr.

For elastic scattering  $(k_i = k_f = 2\pi/\lambda)$ 

 $\vec{Q} = \vec{k}_i - \vec{k}_f$ 

In general, diffraction (SANS or NR) probes length scale

 $d \approx \frac{2\pi}{Q}$ , for small scattering angles,  $d \approx \frac{\lambda}{2\theta}$ 

structure in the direction of Q, on a scale,  $d \approx 2\pi/|{
m \ddot{Q}}|$ More specifically, diffraction (SANS or NR) probes





Reflectivity probes structure perpendicular to surface (parallel to Q), and *averages over structure in plane of sample*.

SANS probes structure in plane of sample (parallel to Q), and averages over structure perpendicular to sample surface.







## Length Scales Probed by SANS and NR

NR probes structure on a scale d, where

 $d \approx \frac{2\pi}{Q} \approx \frac{\lambda}{2\theta}$  (wavelength) (reflection angle)

 $0.4 \text{ nm} < \lambda < 0.6 \text{ nm}$  $0.06^{\circ} < \theta < 20^{\circ} \text{ (small angles)}$ 0.5 nm < d < 500 nm





Length Scales Probed by SANS	s and NR
30-m SANS	Ľ Ľ
$d_{max} = d_{max} \approx \frac{\lambda}{\Delta \theta_{min}} \approx 300_1$ (limited by instrument resolution; in effect, source strength)	$\operatorname{m}  \operatorname{d}_{\max} \approx \frac{\lambda}{\Delta \theta_{\min}} \approx 500 \operatorname{nm}$
$d_{min} d_{min} = \int_{p_{p}}^{1} (0) = \phi V_{p} (\Delta \rho)^{2} d_{s} d_{s}$ $d_{min} = \phi = volume fraction (limited by V_{p}^{p} + 'particle' volume signal-to-noise) \Delta \rho = scattering contrast \Delta \rho = scattering contrast \Delta \rho = scattering contrast \Delta \rho$	$\Gamma_{s}$ $R_{min} \approx 10^{-8} \approx \frac{(8\pi\Delta\rho)^{2}}{Q_{max}^{4}}$ for NCNR instruments
u <sub>s</sub> = sample thickness T <sub>s</sub> = sample transmissior ش ع	for $\Delta \rho \approx 3 \times 10^{-6} \mathrm{A}^{-2}$ (good contrast)
for $V_p = \frac{\sigma}{6} d^3$ and 'good' contrasting particle diameter for NCNR 30 m SANS $-\phi d^3 \ge 5 \times 10^{-4} \text{ nm}^3$	d $d_{max} \sim 0.8 \text{A}^{-1}$ $d_{max} \sim 0.8 \text{nm}$
instruments d <sub>min</sub> φ 1 nm 0.05 %	Qmax
C Center for Neuron Research	

Techniques for the Measurement of Microstructure







- Deuterium Labeling and H- and D-Solvent Mixtures Unique Control of Scattering Contrast by
- Uniquely Powerful Probe of Magnetic Structure

#### Also,

- highly penetrating even at long wavelengths
- equally sensitive to light and heavy elements
- nondestructive



SANS APPLICATIONS	POLYMERS:	<ul> <li>Conformation of Polymer Molecules in Solution <u>and</u> in the bulk</li> <li>Structure of Microphase-Separated Block Copolymers</li> <li>Factors Affecting Miscibility of Polymer Blends</li> </ul>	BIOLOGY:	<ul> <li>Organization of Biomolecular Complexes in Solution</li> </ul>	<ul> <li>Conformation Changes Affecting Function of Proteins, Enzymes, DNA/Protein</li> </ul>	complexes, Membranes, etc.	<ul> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling</li> </ul>	CHEMISTRY:	Structure and Interactions in Colloidal Suspensions, Microemulsions,	Surfactant Micelies, etc.	<ul> <li>Microporosity of Chemical Absorbents</li> </ul>	<ul> <li>Mechanisms of Molecular Self-Assembly in Solutions and on Surfaces of Microporous Media</li> </ul>	Center for Neutron Research	<ul> <li>SANS APPLICATIONS</li> <li>POLYMERS:</li> <li>Conformation of Polymer Molecules in Solution <u>and</u> in the bulk</li> <li>Structure of Microphase-Separated Block Copolymers</li> <li>Eactors Affecting Miscibility of Polymer Blends</li> <li>Factors Affecting Miscibility of Polymer Blends</li> <li>BIOLOGY:</li> <li>Organization of Biomolecular Complexes in Solution</li> <li>Mechanism of Molecular Self-Assembly in Solutions and on Surfaces</li> <li>Microporous Media</li> </ul>
<ul> <li>POLVMERS:</li> <li>Conformation of Polymer Molecules in Solution <u>and</u> in the bulk</li> <li>Euroture of Microphase-Separated Block Copolymers</li> <li>Eactors Affecting Miscibility of Polymer Blends</li> <li>Factors Affecting Miscibility of Polymer Blends</li> <li>BIOLOGY:</li> <li>Organization of Biomolecular Complexes in Solution</li> <li>Organization of Biomolecular Complexes in Solution</li> <li>Conformation Changes Affecting Function of Proteins, Enzymes, DNA/Protein complexes, Membranes, etc.</li> <li>Wechanisms and Pathways for Protein Folding and DNA Supercoiling</li> <li>CHEMISTRY:</li> <li>Structure and Interactions in Colloidal Suspensions, Microemulsions, Surfactant Micelles, etc.</li> <li>Microporous Media</li> <li>Microporous Media</li> </ul>	<ul> <li>Conformation of Polymer Molecules in Solution <u>and</u> in the bulk</li> <li>Structure of Microphase-Separated Block Copolymers</li> <li>Eactors Affecting Miscibility of Polymer Blends</li> <li>Factors Affecting Miscibility of Polymer Blends</li> <li>Organization of Biomolecular Complexes in Solution         <ul> <li>Organization of Biomolecular Complexes in Solution</li> <li>Organization of Biomolecular Complexes in Solution</li> <li>Conformation Changes Affecting Function of Proteins, Enzymes, DNA/Protein complexes, Membranes, etc.</li> <li>Conformation Changes for Protein Folding and DNA Supercoiling CHEMISTRY:</li> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling CHEMISTRY:</li> <li>Mechanisms of Chemical Suspensions, Microemulsions, Surfactant Micelles, etc.</li> </ul> </li> <li>Microporous Media</li> <li>Microporous Media</li> <li>Microporous Media</li> </ul>	<ul> <li>BIOLOGY:</li> <li>Organization of Biomolecular Complexes in Solution</li> <li>Organization of Biomolecular Complexes in Solution</li> <li>Conformation Changes Affecting Function of Proteins, Enzymes, DNA/Protein complexes, Membranes, etc.</li> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling</li> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling</li> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling</li> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling</li> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling</li> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling</li> <li>Mechanisms of Molecular Self-Assembly in Solutions and on Surfaces of Microporous Media</li> </ul>	<ul> <li>Organization of Biomolecular Complexes in Solution</li> <li>Conformation Changes Affecting Function of Proteins, Enzymes, DNA/Protein complexes, Membranes, etc.</li> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling</li> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling</li> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling</li> <li>CHEMISTRY:         <ul> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling</li> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling</li> </ul> </li> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling</li> <li>CHEMISTRY:         <ul> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling</li> <li>Microporosity for Protein Folding and DNA Supercoiling</li> <li>Microporosity of Chemical Suspensions, Microemulsions, Suffactant Microlelles, etc.</li> <li>Microporosity of Chemical Absorbents</li> <li>Microporous Media</li> </ul> </li> <li>Mater Internetions for Molecular Self-Assembly in Solutions and on Surfaces of Microporous Media</li> </ul>	<ul> <li>Conformation Changes Affecting Function of Proteins, Enzymes, DNA/Protein complexes, Membranes, etc.</li> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling</li> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling</li> <li>Structure and Interactions in Colloidal Suspensions, Microemulsions, Surfactant Micelles, etc.</li> <li>Microporosity of Chemical Absorbents</li> <li>Microporous Media</li> </ul>	<ul> <li>complexes, Membranes, etc.</li> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling</li> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling</li> <li>Structure and Interactions in Colloidal Suspensions, Microemulsions, Surfactant Micelles, etc.</li> <li>Microporosity of Chemical Absorbents</li> <li>Microporous Molecular Self-Assembly in Solutions and on Surfaces of Microporous Media</li> </ul>	<ul> <li>Mechanisms and Pathways for Protein Folding and DNA Supercoiling CHEMISTRY:</li> <li>Structure and Interactions in Colloidal Suspensions, Microemulsions, Surfactant Micelles, etc.</li> <li>Microporosity of Chemical Absorbents</li> <li>Microporosity of Chemical Absorbents</li> <li>Microporous Molecular Self-Assembly in Solutions and on Surfaces of Microporous Media</li> </ul>	CHEMISTRY: • Structure and Interactions in Colloidal Suspensions, Microemulsions, Surfactant Micelles, etc. • Microporosity of Chemical Absorbents • Mechanisms of Molecular Self-Assembly in Solutions and on Surfaces of Microporous Media	<ul> <li>Structure and Interactions in Colloidal Suspensions, Microemulsions, Surfactant Micelles, etc.</li> <li>Microporosity of Chemical Absorbents</li> <li>Mechanisms of Molecular Self-Assembly in Solutions and on Surfaces of Microporous Media</li> </ul>	A Microporosity of Chemical Absorbents     Microporosity of Chemical Absorbents     Mechanisms of Molecular Self-Assembly in Solutions and on Surfaces     of Microporous Media	<ul> <li>Microporosity of Chemical Absorbents</li> <li>Mechanisms of Molecular Self-Assembly in Solutions and on Surfaces of Microporous Media</li> </ul>	Mechanisms of Molecular Self-Assembly in Solutions and on Surfaces     of Microporous Media	Center for Neutron Research		

In Situ SANS Study of Evolution of Porosity in Low-K Films	C. Kim, K. Shin nd R.D. Miller SUNY Stonybrook 1 Res. Center	deuterated poragen to introduce contrast in as-spun films ow evolution of poragen kfill poragen-free final material with deuterated wetting I to compare pore with poragen structure $100 \int_{10}^{10} \int_{125^{\circ}C} \int_{10}^{125^{\circ}C} \int_{10}^{125^{\circ}C} \int_{10}^{10} \int$	SINCHAL
	R.M. Briber and G.Y. Yang E. Huang, H. University of Maryland W. Volksen a IBM Almade	<ul> <li>I(q) cm<sup>1</sup></li> <li>I(q) cm<sup>1</sup></li> </ul>	Center for Neutron Research

### METALS AND CERAMICS:

- Kinetics and Morphology of Precipitate Growth in Alloys and Glasses
- Defect Structures (e.g. microcracks, voids) Resulting from Creep, Fatigue or Radiation Damage
- Grain and Defect Structures in Nanocrystalline Metals and Ceramics

#### MAGNETISM:

- Magnetic Ordering and Phase Transitions in Ferromagnets, Spin Glasses, Magnetic Superconductors, etc.
- Flux-Line Lattices in Superconductors





# Vortex Matter in Superconductor Nb









#### Polymers:

- polymer phase behavior in thin films
- e.g. order-disorder phase transitions of block copolymers





#### Polymers:

- Polymer Interdiffusion at Interfaces
- Factors Affecting Wetting and Dewetting
- Polymer conformation and concentration profiles at solid-liquid and liquid-air interfaces

#### Chemistry:

- Langmuir-Blodgett films
- Self-Assembled monolayers, bilayers, etc.
- Electrochemical reactions





# NR Study of Surfactants at Electrode Surfaces

I. Burgess, et al. (U. Guelph), J. Majewski & G. Smith (LANL), S. Satija & R. Ivkov (NCNR)



**SGS** Center for Neutron Research

Length [Ă]

PIN V VII

#### **Biology:**

- Location of Peptides in Biomimetic Single Bilayer Membranes
- Protein Adsorption/Desorption on Self-Assembled Monolayers
- Vectorially-Oriented Protein Monolayers

#### Magnetism:

- Magnetic Structure and Interlayer Coupling in GMR Multilayer Thin Films
- Magnetic Coupling and Ordering across Non-Magnetic Layers
- Spin Structures Associated with Exchange-Biased Magnetic Thin Films









### Therefore, diffraction probes structure in the direction of Q only!

O, divided by # incident per direction corresponding to Scattering cross section: # neutrons scattering in unit area

 $\frac{\mathrm{d}\sigma}{\mathrm{d}\Omega} = \frac{1}{N} \sum_{i} b_{i} e^{i \vec{\mathrm{Q}} \cdot \vec{\mathrm{I}}_{i}}$ 

Appendix A. Scattering from N Nuclei



**Only components** 

of r<sub>i</sub> parallel to Q

contribute to

summation

 $\mathbf{Q} \cdot \vec{\mathbf{r}} = \mathbf{Q} \mathbf{r}_{//}$  $\mathbf{I}_{\mathbf{I}}^{\mathsf{T}} = \mathbf{I}_{\mathbf{I}}^{\mathsf{T}} + \mathbf{I}_{\mathbf{I}}^{\mathsf{T}}$